

Green aldose isomerisation: 2-*C*-methyl-1,4-lactones from the reaction of Amadori ketoses with calcium hydroxide

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Abstract—Saccharinic acids, branched 2-*C*-methyl-aldonic acids, may be accessed via a green procedure from aldoses by sequential conversion to an Amadori ketose and treatment with calcium hydroxide; *D*-galactose and *D*-glucose are converted to 2-*C*-methyl-*D*-lyxono-1,4-lactone (with a small amount of 2-*C*-methyl-*D*-xylono-1,4-lactone) and 2-*C*-methyl-*D*-ribono-1,4-lactone. Inversion of configuration at C-4 of the branched lactones allows access to 2-*C*-methyl-*L*-ribono-1,4-lactone and 2-*C*-methyl-*L*-lyxono-1,4-lactone, respectively. *D*-Xylose affords 2-*C*-methyl-*D*-threono-1,4-lactone and 2-*C*-methyl-*D*-erythrono-1,4-lactone, whereas *L*-arabinose, under similar conditions, gave the enantiomers 2-*C*-methyl-*L*-threono-1,4-lactone and 2-*C*-methyl-*L*-erythrono-1,4-lactone.

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At present, all commercially available carbohydrate scaffolds contain linear carbon chains.¹ The Kiliani cyanide ascensions^{2,3} on unprotected ketohexoses—*D*-fructose, *L*-sorbose,⁴ *D*-tagatose and *D*-psicose⁵—provide an accessible set of derivatives bearing a C-2 hydroxymethyl group of value in the efficient synthesis of complex homochiral targets;⁶ the Kiliani reaction of the C-2 branched sugars^{7,8} gives C-3 branched sugars. The isomerisation of aldoses or ketoses into 2-*C*-methyl-aldonic acids by calcium hydroxide in very low yields is one of the oldest⁹ and most complex organic reactions with typically less than 0.5% of branched 2-*C*-methyl-aldonic acids being formed;¹⁰ after optimisation of the reaction of fructose with calcium hydroxide a yield of around 11% of 2-*C*-methyl-*D*-ribonic acid **3D** can be obtained in several weeks.¹¹ This letter establishes the generality of the calcium hydroxide isomerisations of Amadori ketoses, derived from cheap aldoses, to 2-*C*-methyl-branched aldonic acids as a new strategy for the synthesis of branched sugar chirons.

The sole previous report of this transformation (Scheme 1) is the conversion of *D*-glucose **1** with dimethylamine to fructosamine **2**, which with aqueous calcium hydroxide

gives the branched ribose derivative **3D**, isolated as its lactone;¹² a practical procedure is given below for the transformation of *D*-galactose **6** via the Amadori ketose **5** to 2-*C*-methyl-lyxonic acid **4D**. Although the yields are modest, the low cost of glucose and galactose make **3D** and **4D** accessible starting materials for homochiral targets with methyl-branched carbon chains. There are currently no cheaply available *L*-aldohexoses; however, inversion of configuration at C-4 of 1,4-lactones can be accomplished efficiently on a multi-kilogram scale.¹³ Thus the epimerisations of *D*-ribono-**3D** to *L*-lyxono-**4L** and of *D*-lyxono-**4D** to *L*-ribono-**3L** allow both enantiomers to be synthesised.

Pentoses also undergo the same transformation (Scheme 2). Thus, *D*-xylose **7** was converted to Amadori derivative **8**; subsequent reaction of **8** with calcium oxide allows the isolation of both 2-*C*-methyl-*D*-erythrono-**9D** and *D*-threono-**10D** acids. *L*-Arabinose **12**, the only readily available *L*-pentose, allows the formation of enantiomers **9L** and **10L** from *L*-ribulosamine **11**.

D-Galactose **6** on reaction with dibenzylamine in acetic acid/ethanol (Scheme 3) underwent the Amadori rearrangement to give tagatosamine **5**, which crystallised as the α -anomer,¹⁴ in 88% yield.¹⁵ Treatment of Amadori ketose **5** with calcium oxide in water, followed

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A practical method for the synthesis of branched D-lyxonolactone **13D** (the lactone derived from **4D**) is as follows: A suspension of the Amadori ketose **5** (41.2 g, 115 mmol) in water (750 mL) was stirred at 70 °C with calcium oxide (32 g, 570 mmol) for 24 h. The reaction mixture was cooled to room temperature, filtered through Celite® and then passed through Amberlite® IR-120 H⁺ ion exchange resin, using water as the eluant. The water was then removed under reduced pressure to afford a crude orange oil which was purified by flash column chromatography (ethyl acetate/cyclohexane, (6:1) → ethyl acetate) to afford 2-C-methyl-D-lyxono-1,4-lactone **13D** (1.98 g, 12%)¹⁶ and D-xylono-epimer **14** (338 mg, 2%).¹⁷ The structure of the unprotected lyxono-lactone **13D** was confirmed by X-ray crystallographic analysis;¹⁸ reaction of xylono-epimer **14** with acetone in the presence of anhydrous copper sulfate and acid gave 3,5-acetonide **15** (mp 155–158 °C, $[\alpha]_D^{23} +82.2$ (*c* 0.67, CHCl₃)), an X-ray structure of which was also determined.¹⁹ It is easy to distinguish between the diastereomers by their ¹³C spectra; the chemical shift of the 2-C-methyl group of lyxono-isomer **13D** (δ 20.7) where the diol unit is *cis* is at a significantly lower field than xylono-isomer **14** (δ 16.8), where the diol unit is *trans*.

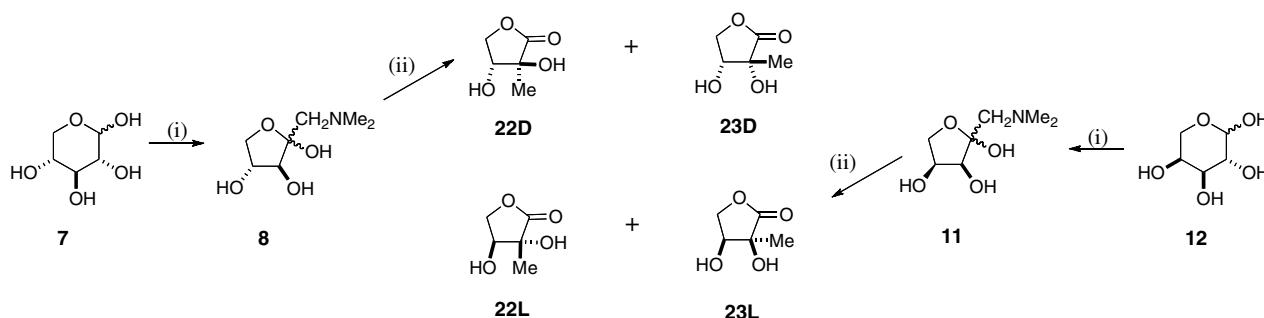
The branched D-lyxono-lactone **13D** was converted into L-ribo- lactone **19L**, epimeric at C-4. Acetona- tion of **13D** gave the 5 ring 2,3-O-isopropylidene derivative **16D** (mp 64–66 °C, $[\alpha]_D^{23} +77.2$ (*c* 1.75, Me₂CO)) in 99% yield; in contrast to the reaction of lyxonolactone itself,²⁰ none of the 6 ring 3,5-ketal was formed. Esterification of the free alcohol in **16D** with triflic anhydride in dichloromethane in the presence of pyridine gave triflate **17D** (mp 79–81 °C, $[\alpha]_D^{21} +60.9$ (*c* 0.60, Me₂CO)) in 100% yield. Reaction of the triflate **17D** with potassium hydroxide in aqueous dioxane caused ring opening and subsequent epoxide formation; work-up with acid gave inversion of configuration at C-4 and closure to the protected lactone **18L** (mp 52–54 °C, $[\alpha]_D^{22} +34.2$ (*c* 1.05, Me₂CO)) in 82% yield. Removal of the ketal from **18L** with aqueous trifluoroacetic acid gave the unprotected L-ribo- lactone **19L** (mp 160–162 °C, $[\alpha]_D^{22} -87.7$ (*c* 0.60, water) {lit.¹¹ for the D-enantiomer **19D** mp 160–161 °C, $[\alpha]_D^{22} +93$ (water)}) in 90% yield. The overall yield for the conversion of D-lyxono-lactone **13D** (the lactone of **4D**) to L-ribo- lactone **19L** (the lactone of **3L**) was 73%.

A similarly efficient epimerisation of D-ribo- lactone **19D** (equivalent to the open chain aldonic acid **3D**) into L-lyxono-lactone **13L** (equivalent to the aldonic acid **4L**) was achieved in 68% overall yield. Lactone **19D**, prepared in 20% yield from D-glucose **1**,¹² was protected as its 2,3-acetonide **18D** (mp 52–54 °C; $[\alpha]_D^{21} -35.6$ (*c* 1.42, Me₂CO)) in 100% yield. Reaction of **18D** with mesyl chloride in pyridine afforded mesylate **21D** (oil, $[\alpha]_D^{24} -24.7$ (*c* 0.97, CHCl₃)) in 100% yield. Treatment of **21D** with potassium hydroxide in aqueous dioxane followed by an acid work-up gave inversion at C-4 to give the protected lactone **16L** (mp 64–66 °C, $[\alpha]_D^{22} -74.1$ (*c* 0.64, Me₂CO), 91% yield) which was deprotected by aqueous trifluoroacetic acid to afford the branched L-lyxono-lactone **13L** (oil, $[\alpha]_D^{18} -77.3$ (*c* 0.4, water)) in 75% yield.

2-C-Methyl-D-erythrono-1,4-lactone **23D**, the lactone of **9D**, was discovered as a natural product in *Astragalus lusitanicus*²¹ and *Cicer arietinum*;²² D-threono-epimer **22D** was isolated from *Anthyllis tetraphylla*²³ and also found in tobacco smoke.²⁴ 2-C-Methyl-D-erythritol was discovered in the petals of *Phlox subulata* and is involved in flower development.²⁵ The mevalonate-independent biosynthesis of isoprenoids via 2-C-methyl-D-erythritol phosphate is a target against malaria and pathogenic bacteria²⁶ and has increased the effort in the synthesis of 2-C-methyl tetroses.²⁷

Previous syntheses of lactones **22D** and **23D**—all of which involve several steps and need protection of oxygen functional groups—include asymmetric aldol condensations^{28,29} and use of protected chiral pool starting materials of lactic acid³⁰ and mannitol.³¹ Although very small amounts of D-lactones **22D** and **23D** are obtained by the reaction of calcium hydroxide with D-xylose,³² the sequential Amadori rearrangement-calcium hydroxide treatment of the pentoses D-xylose **7** and L-arabinose **12** produced enantiomeric lactones **22D** and **23D** or **22L** and **23L**, respectively, without any protection necessary.

Thus, D-xylose **7** was subjected to the Amadori rearrangement by treatment with ethanolic dimethylamine in acetic acid to give the xylulosamine **8** (Scheme 4); crude residue **8** obtained by removal of the solvent was treated with calcium oxide. After workup with acid ion exchange resin, a mixture of the branched D-threono-**22D** (10%)³³ and D-erythrono-**23D** (4%)³⁴ lactones was



Scheme 4. Reagents: (i) Me₂NH, AcOH, EtOH; (ii) CaO, H₂O; then H⁺.

obtained. Neither of the lactones was crystalline; very careful chromatography was necessary to obtain pure samples of each separate diastereomer. The enantiomeric branched *L*-threono-**22L** and *L*-erythro-**23L** lactones were obtained using an identical procedure from *L*-arabinose **12** via the intermediate ribulosamine **11**; the ratio of *L*-threono-**22L**/*L*-erythro-**23L** was also approximately 3:1. Again the diastereomers could be differentiated by ^{13}C spectroscopy; the chemical shift of the 2-*C*-methyl group of the *cis*-diol erythro-*isomer* **23** (δ 19.9) is at significantly lower field than the *trans*-diol threono-*isomer* **22** (δ 16.6).

It is noteworthy that the products from the aldohexoses (glucose and galactose) have a 2,3-*cis*-diol in the lactone ring as the major products, whereas the aldopentoses (xylose and arabinose) form predominantly the lactone with a *trans*-diol.

In summary, the isomerisation of aldoses to saccharinic (2-*C*-methyl-aldonic) acids by an Amadori rearrangement followed by treatment with aqueous calcium oxide is shown to be general and provides access by green aqueous procedures to a group of hitherto unavailable branched carbohydrate chirons.

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- 2-*C*-Methyl-*D*-lyxono-1,4-lactone **13D**: mp 106–107 °C; $[\alpha]_{\text{D}}^{23}$ +70.4 (*c* 0.9, Me₂CO) ν_{max} (film): 3421, 1773 cm⁻¹; δ_{H} (CD₃OD, 400 MHz): 1.43 (3H, s, CH₃), 3.88 (2H, m, H-5 and H-5'), 4.07 (1H, d, *J*_{3,4} 3.8 Hz, H-3), 4.54 (1H, ddd, *J*_{4,5} 5.0 Hz, *J*_{3,4} 3.8 Hz, *J*_{4,5'} 5.9 Hz, H-4); δ_{C} (CD₃OD, 100.6 MHz): 20.7 (CH₃), 60.4 (C-5), 74.2 (C-3), 74.5 (C-2), 81.5 (C-4), 179.4 (C-1).
- 2-*C*-Methyl-*D*-xylono-1,4-lactone **14**: mp 161–162 °C; $[\alpha]_{\text{D}}^{23}$ +87.3 (*c* 0.5 in water); ν_{max} (film): 3254, 1776 cm⁻¹; δ_{H} (CD₃OD, 400 MHz): 1.39 (3H, s, CH₃), 3.82–3.90 (2H, m, H-5 and H-5'), 4.03 (1H, d, *J*_{3,4} 3.7 Hz, H-3), 4.71 (1H, ddd, *J*_{3,4} 3.7 Hz, *J*_{4,5} 5.0 Hz, *J*_{4,5'} 6.4 Hz, H-4); δ_{C} (CD₃OD, 100.6 MHz): 16.8 (CH₃), 60.4 (C-5), 75.3 (C-3), 76.5 (C-2), 82.9 (C-4), 178.2 (C-1).
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- 2-*C*-Methyl-*D*-threono-1,4-lactone **22D**: oil, $[\alpha]_{\text{D}}^{23}$ –18.8 (*c* 1.1, H₂O); ν_{max} (film): 3406, 1775 cm⁻¹; δ_{H} (CD₃OD, 400 MHz): 1.36 (3H, s, CH₃), 3.98 (1H, dd, *J*_{3,4} 4.4 Hz, *J*_{4,4'} 9.4 Hz, H-4), 4.19 (1H, m, H-3), 4.50 (1H, dd, *J*_{3,4'} 5.4 Hz, *J*_{4,4'} 9.4 Hz, H-4'); δ_{C} (CD₃OD, 100.6 MHz): 16.6 (CH₃), 71.8 (C-4), 74.5 (C-3), 74.9 (C-2), 179.0 (C-1).
- 2-*C*-Methyl-*D*-erythro-1,4-lactone **23D**: oil, $[\alpha]_{\text{D}}^{22}$ –35.0 (*c* 0.3, H₂O); ν_{max} (film): 3398, 1774 cm⁻¹; δ_{H} (CD₃OD, 400 MHz): 1.41 (3H, s, CH₃), 4.06 (1H, dd, *J*_{3,4} 1.8 Hz, *J*_{3,4'} 4.1 Hz, H-3), 4.16 (1H, dd, *J*_{3,4} 1.85 Hz, *J*_{4,4'} 10.2 Hz, H-4), 4.45 (1H, dd, *J*_{3,4'} 4.1 Hz, *J*_{4,4'} 10.2 Hz, H-4'); δ_{C} (CD₃OD, 100.6 MHz): 19.9 (CH₃), 72.3 (C-4), 73.4 (C-2), 73.5 (C-3), 179.4 (C-1).